NucleoSpin® RNA Virus

1 Kit contents

	NucleoSpin® RNA Virus	
REF	**12 preps 740956.12C**	**240 preps 740956.240C**
Lysis Buffer RAV1	2 x 10 mL	2 x 120 mL
Wash Buffer RAW	2 x 6 mL	3 x 75 mL
Wash Buffer RAV3 (Concentrate)	6 mL (add 24 mL ethanol before first use)	2 x 25 mL (add 100 mL ethanol to each bottle before first use)
Elution Buffer RE	30 mL	2 x 30 mL
Carrier RNA (lyophilized)	300 µg	10 x 300 µg
NucleoSpin® RNA Virus Columns (dark blue rings, plus Collection Tubes)	12	4 x 60
User manual	1	1

2 How to use the kit

Please see the protocol information how to use the kit (see next pages). For further questions and more detailed information, please contact MACHEREY-NAGEL at tech-bio@mn-net.com for protocol information how to use the kit on specific robotic instruments.

For storage conditions, product use restrictions, and safety information, please see the general NucleoSpin® RNA Virus user manual.

NucleoSpin® kits on QIAcube®
MN is not recommending to use this kit on specific robots. The use of NucleoSpin® kits on theQIAcube® is solely at your own discretion. MACHEREY-NAGEL is not responsible for loss of warranty claims or other consequences.

3 General information

Application:	Virus
Kit:	NucleoSpin® RNA Virus (REF 740956.240C) instead of: QIAamp® Viral RNA Mini Kit
Sample material:	Body fluid
Protocol name:	Purification of viral RNA from cell-free body fluids Virus_QIAampViralRNA_BodyFluid_Standard_V2
Editable parameters:	Elution volume: 50–100 µL; default 60 µL

4 Using the kit

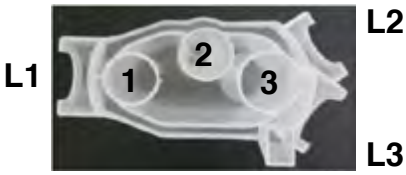
1. Fill the designated buffer bottles with the buffers according to the buffer table below.
2. Samples: 140 µL cell-free body fluids
3. Place Sample Tubes (2 mL screw-cap tubes without skirted base) into sample rack (shaker).
4. Reconstitute lyophilized Carrier RNA (each 300 µg vial; suitable for 24 runs) in 300 µL Buffer RAV1. Dilute Carrier RNA according to the table below.
5. Insert disposable Filter Tips 1000 µL.
6. General equipment setup is shown below.

5 Additional materials

Refer to the QIAcube® protocol sheet for required consumables (e.g., sample tubes, collection tubes, instrument accessories, disposable tips, etc.) and software requirements.

6 Rotor adapter

Position	Labware	Lid position
1	NucleoSpin® RNA Virus Column	L1
2	–	–
3	1.5 mL collection tube*	L3



* Sarstedt, Micro tube 1.5 mL Safety Cap

7 Buffers (Reagent Bottle Rack)

Position	MN Reagent	Replaced Reagent QIAGEN®
1	Buffer RAV1	Buffer AVL
2	100 % ethanol	100 % ethanol
3	Buffer RAW	Buffer AW1
4	Buffer RAV3	Buffer AW2
5	–	–
6	–	–

8 Microcentrifuge Tube Slots

	Position A	Position B	Position C
Content:	–	Buffer RE	Diluted Carrier RNA
Tube:	–	2 mL screw-cap tube without skirted base	2 mL screw-cap tube without skirted base

9 Required volume of Elution Buffer and diluted Carrier RNA in Microcentrifuge Tube Slots

No. of samples	Elution buffer (Microcentrifuge Tube Slot B)	Diluted Carrier RNA (Microcentrifuge Tube Slot C)
2	256 µL	125 µL (28 µL Carrier RNA + 97 µL Buffer RE)
3	364 µL	150 µL (33.6 µL Carrier RNA + 116.4 µL Buffer RE)
4	472 µL	175 µL (39.2 µL Carrier RNA + 135.8 µL Buffer RE)
5	580 µL	200 µL (44.8 µL Carrier RNA + 155.2 µL Buffer RE)
6	688 µL	225 µL (50.4 µL Carrier RNA + 174.6 µL Buffer RE)
7	796 µL	250 µL (56 µL Carrier RNA + 194 µL Buffer RE)
8	904 µL	275 µL (61.6 µL Carrier RNA + 213.4 µL Buffer RE)
9	1012 µL	300 µL (67.2 µL Carrier RNA + 232.8 µL Buffer RE)
10	1120 µL	325 µL (72.8 µL Carrier RNA + 252.2 µL Buffer RE)
12	1336 µL	375 µL (84 µL Carrier RNA + 291 µL Buffer RE)

10 Internal control

If an internal control is to be used, it can be supplied in position C. For each prepared sample (plus 3 additional samples) supply the appropriate amount in 25 µL together with the Carrier RNA according to the table below.

No internal control	With internal control
Carrier RNA: 5.6 µL	Carrier RNA: 5.6 µL
Buffer RAV1: 19.4 µL	Internal control: Up to 19.4 µL (if < 19.4 µL internal control is used, adjust final volume to 25 µL with Buffer RAV1)

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